

# PROLINE TUTORIAL

## QUANTIFY BY SPECTRAL COUNTING

### // START PROLINE

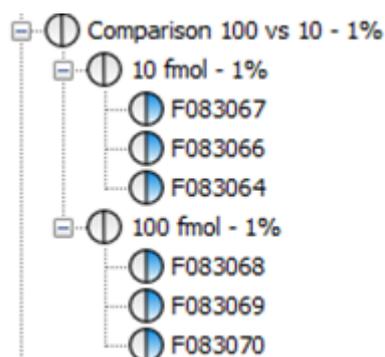
#### A. EXPERIMENTAL DESIGN

In this tutorial, all sample mascot files available on Proline website will be used. This dataset contains the MS analysis of two samples of 2 µg yeast cell lysate spiked respectively with 100fmol and 10fmol of UPS1. Samples were analyzed in triplicate by nanoLC–MS/MS on an LTQ–Orbitrap Velos mass spectrometer. For more information on samples preparation and LC–MS/MS analyses, please refer to Ramus et al., J Proteomics. 2016 Jan 30;132:51-62. doi: 10.1016/j.jprot.2015.11.011.

#### Action

Reproduce the following experimental design

- Create all datasets
- Import of the missing Mascot files (see Proline Basics tutorial)
- If necessary, rename the files according to the Search Result Name to reproduce the experimental design below:



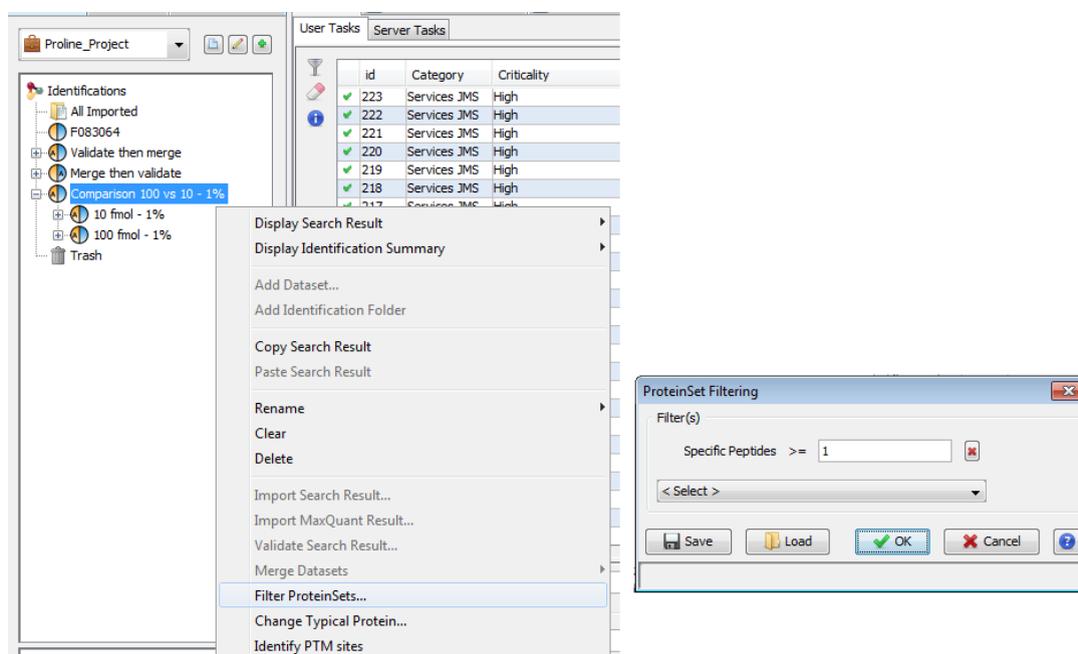
#### Note

*Files and datasets can be renamed manually. Files can also be renamed automatically by retrieving the Search Result name for example*

**Comparison 100 vs 10 1%** is a merge by aggregation of identification summaries created at the intermediate level: 10 fmol - 1% and 100 fmol - 1%. These both datasets are also merge by aggregation of identification summaries (from imported result files), validated with the following parameters : 1% PSM (based on score) and rank = 1. The top level dataset ("Comparison 100 vs 10 - 1%) has been filtered to retain only protein sets with at least 1 specific peptide.

### Action

- Validate the six search results using the described parameters. Note: Validation can be done on multiple search results.
- Merge the resulting identification summary at intermediate and top level of the dataset hierarchy.
- Filter the top level identification summary to retain only protein sets with at least 1 specific peptide.

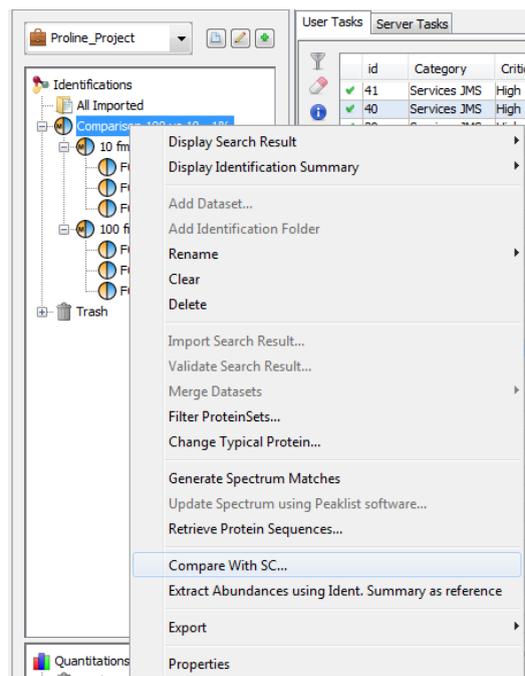


## II/ COMPARE SAMPLES BY SPECTRAL COUNTING

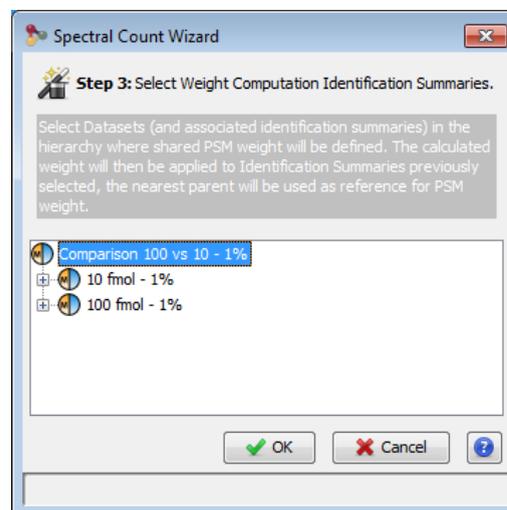
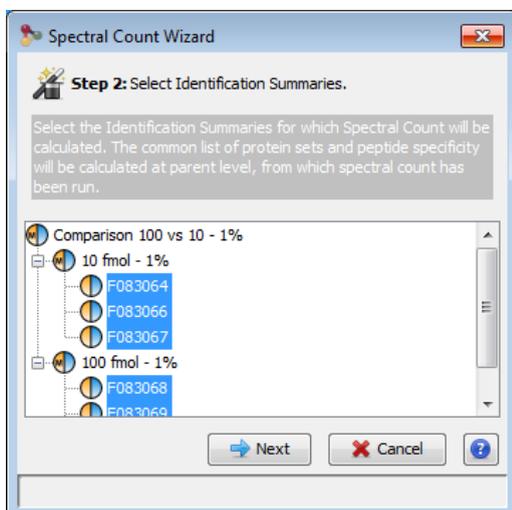
### A. RUN SPECTRAL COUNTING

To run the SC comparison, you should have a merged dataset containing child datasets (which may be also merged datasets or identification datasets). Actually, only identification summaries merge could be used to execute SC.

To execute SC comparison, right-click on merged dataset and select *Compare with SC*.



The opened dialog allows you to specify a name and a description for the comparison. On the second dialog box, Step 2, select the dataset on which you would like to perform the Spectral Count (in our case, we would like to compute the SC value for each of the six datasets) and finally choose the dataset where shared peptides spectral count weights will be calculated (in our case, the top level dataset).



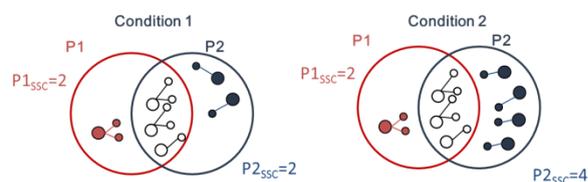
## B. COMPUTED VALUES

The algorithm implemented in Proline compute three different spectral count values :

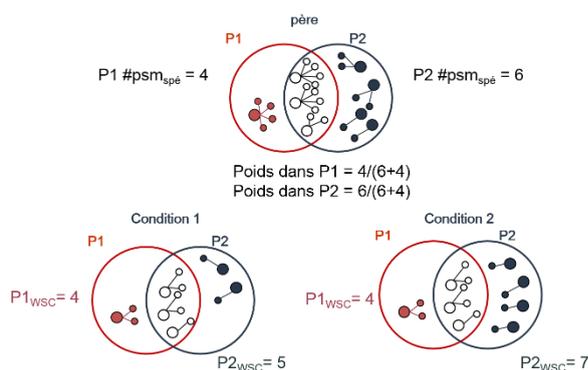
1/ **Basic SC**: is the total number of MS/MS validated peptide spectrum match (PSM) of all peptides matching the considered protein set.



2/ **Specific SC**: Is the total number of validated PSM of peptides that are **specific** to the considered protein set. Shared peptides (peptides shared by different protein sets) are excluded from the count. The peptide specificity is calculated from the top level identification summary to ensure that the uniqueness of the protein set the peptide belongs to is not modified by an additional peptide identification from another result summary.

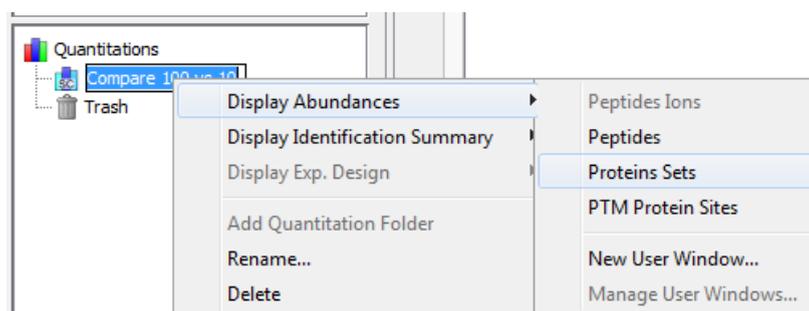


3/ **Weighted SC**: is based on all identified peptides, but taking into account that spectral count of shared peptides must be apportioned to reflect the contribution of each protein set. The weight of these respective contributions is based on the number of specific peptides of each considered protein set at the top level identification summary.



### C. RESULT VISUALIZATION

Once finished, a new dataset appears in the “Quantitations” panel (lower part of the left window). To visualize SC results, right-click on this dataset and select *Display Abundances* then *Proteins Sets*.



For each replicate, the table columns indicate:  
protein status/ Peptides Count / Basic SC / Specific SC /  
Weighted SC

Protein Set	Overview	#Peptide	#Quant. Peptide	Status F082064	Peptides Count F082064	Basic SC F082064	Specific SC F082064	Weighted SC F082064	Status F082066	Peptides Count F082066	Basic SC F082066
1	HP12_YEAST	41	41	typical	41	122	122	122.00	typical	41	122
2	KPK1_YEAST	46	46	typical	46	132	132	132.00	typical	46	138
3	KPK1_YEAST	43	43	typical	43	121	121	121.00	typical	43	146
4	END2_YEAST	32	32	typical	32	94	94	94.29	typical	32	406
5	GFP3_YEAST	35	35	typical	35	126	113	114.51	typical	35	134
6	PDC1_YEAST	35	35	typical	32	145	145	145.00	typical	33	152
7	HGP2_YEAST	39	39	typical	27	85	28	31.09	typical	26	69
8	HGP2_YEAST	39	39	typical	29	87	46	72.91	typical	30	92
9	HGP7_YEAST	36	36	typical	28	120	28	88.28	typical	31	117
10	TRFL_HUMAN_LIPS	31	31	typical	3	4	4	4.00	typical	4	13
11	HGP72_YEAST	34	34	typical	26	127	27	74.74	typical	30	111

### Note

Every table in Proline can be customized by clicking on the  icon to select visible and invisible columns.

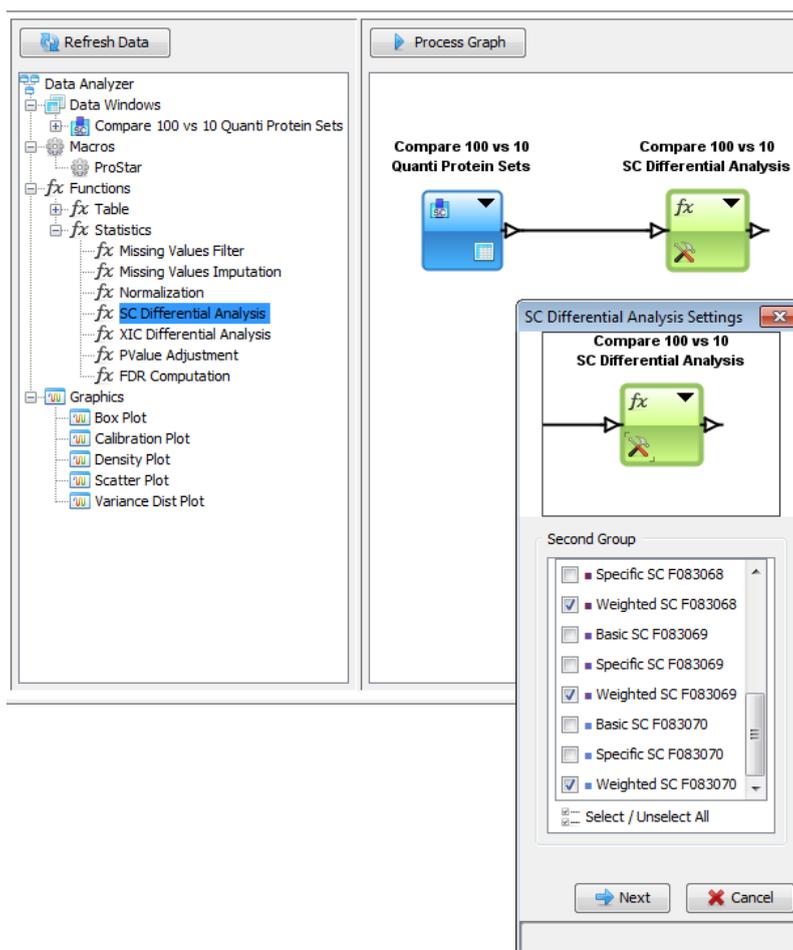
## D. STATISTICAL ANALYSIS

In this section, we will perform the statistical beta binomial test <sup>1</sup> on the **weighted spectral count** data and display the test results.

- (1) Pham, T. V., Piersma, S. R., Warmoes, M., and Jimenez, C. R. (2010) On the beta-binomial model for analysis of spectral count data in label-free tandem mass spectrometry-based proteomics. *Bioinformatics* 26, 363–369

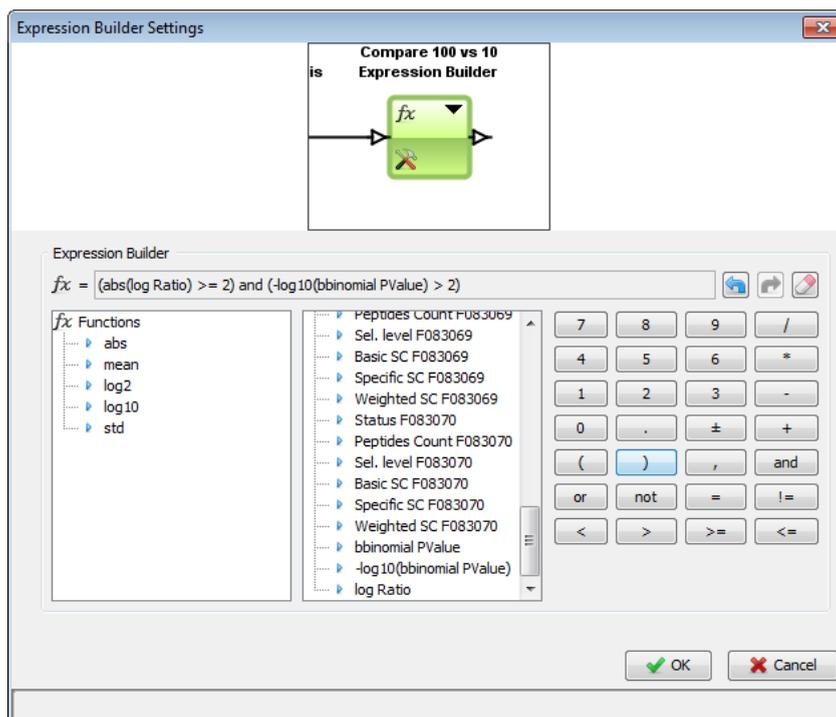
### Action

- Open *Display Abundances* then *Proteins Sets* and click on 
- The DataAnalyser window opens and a box indicating «Compare 100 vs 10 Quanti Protein Sets » appears on the right side of the window.
- Add the « SC Differential Analysis » function to the workflow from Functions > Statistics in DataAnalyser tree, (drag & drop or double click the function) and connect the two boxes.
- Run the statistical function, select the columns belonging to each group to be compared



### Action

- To easily identify proteins of interest, add a column to the table with the « Expression Builder » (from Functions > Table in DataAnalyser tree). Connect the output of SC Differential Analysis box to the entry of the Expression Bulder box. Expression of the new column: proteins with log ratio  $\leq -2$  or  $\geq 2$  and pvalue  $< 0.01$  (use abs() function as shown in the screenshot below)

**Action**

- Visualize results as a Volcano plot: add a graphical view (using  icon) to the newly computed table, select scatter plot and choose log Ratio as x axis and  $-\log_{10}(\text{bbinomial pvalue})$  as y axis.
- In the table, select rows (protein sets) with a non null value in the column that have been added with the « Expression Builder » (use filter button) and visualize those protein sets in the scatter plot.

**Note**

*In Proline, selection can be “transferred” from a view to another view by using the  icon. In the plot, right click on the selected points and create a group containing these points.*

Bravo ! 49 proteins setshave been identified as differentially expressed by the spectral counting approach and the beta binomial statistical test. Among this 49 proteins, 43 proteins out of the 48 UPS1.

